

CURRICULUM VITAE
D. Noah Sather, PhD

CONTACT INFORMATION

D. Noah Sather, PhD
Global, Environmental, and Genomic Health Sciences
Institute for Translational Virology and Innovation
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1. PERSONAL DATA

Legal Name: David Noah Sather, PhD
Place of Birth: Provo, Utah – USA
Citizenship: USA

2. EDUCATION

08/1998 – 07/2002 BS, Biological Sciences, Wayne State University; Detroit, MI
09/2002 – 06/2007 PhD, Biological Sciences, Wayne State University; Detroit, MI

3. POSTGRADUATE TRAINING

07/2007 – 05/2009 Postdoctoral Researcher, Seattle Biomedical Research Institute, Seattle, WA
05/2009 – 02/2011 Senior Scientist, Seattle Biomedical Research Institute, Seattle, WA
02/2011 – 03/2015 Principal Scientist, Seattle Biomedical Research Institute, Seattle, WA

4. FACULTY POSITIONS HELD

03/2015 – 01/2020 Assistant Professor, Center for Global Infectious Disease Research, Seattle Children's Research Institute, Seattle, WA
09/2018 – 01/2020 Affiliate Assistant Professor; Department of Global Health, Pathobiology Program, University of Washington, Seattle, WA
02/2020 – 06/30/2024 Associate Professor; Department of Pediatrics, Division of Infectious Diseases, University of Washington, Seattle, WA
02/2020 – 06/30/2024 Adjunct Associate Professor; Department of Global Health, Pathobiology Program, University of Washington, Seattle, WA

07/2024 – 02/2026	Professor, Department of Pediatrics, Division of Infectious Diseases, University of Washington, Seattle WA
07/2024 – 02/2026	Adjunct Professor, Department of Global Health, Pathobiology Program, University of Washington, Seattle, WA
02/2026 – Present	Professor, Department of Global, Environmental, and Genomic Health Sciences, University of South Florida, Tampa, FL
02/16/2026 - Present	Professor, Center for Global Health and Inter-Disciplinary Research, University of South Florida, Tampa, FL
02/16/2026 - Present	Professor, Institute for Translational Virology and Innovation, University of South Florida, Tampa, FL

5. HOSPITAL POSITIONS HELD

None

6. HONORS

06/2003	Graduate Teaching Award, Biological Sciences, Wayne State University
06/2004	Graduate Teaching Award, Biological Sciences, Wayne State University
08/2004	Dr. William A. Turner Memorial Scholarship, Wayne State University
06/2005	Graduate Teaching Award, Biological Sciences, Wayne State University
06/2006	Summer Dissertation Fellowship, Wayne State University Graduate School
08/2006 – 05/2007	Thomas C. Rumble Competitive Fellow, Wayne State University
02/2009	Keystone Symposium Travel Scholarship, Keystone Symposium, Colorado
02/2009	International Travel Scholarship, AIDS Vaccine 2009, Paris, France

7. BOARD CERTIFICATION

None

8. CURRENT LICENSE(S) TO PRACTICE

None

9. PROFESSIONAL ORGANIZATIONS

None

10. EDUCATION AND TRAINING ACTIVITIES

(a) TABLE: Didactic Teaching – UW

Dates	Course & Title	Responsibility (% if shared course)
01/2019 – 03/2022	PABIO 553: Survival Skills	Co-Instructor- 50%
05/2019 – 05/2020	PABIO 582: Critical Thinking and Research Design	Guest Instructor- 10%
01/2021 – 03/2025	PABIO 580: Pathobiology Seminar	Instructor- 100%
09/2022 – 12/2024	PABIO 551: Biochemistry/Genetic of Pathogens and Their Hosts	Instructor- 100%

(b) TABLE: Thesis Committee Membership

Dates	Student Name	Degree Program	Committee Role
05/2021 – 07/2024	Kristina Edwards	UW Pathobiology	Member
10/2022 – 12/2025	Crystal Chhan	UW Pathobiology	Member
09/2023 – 02/2026	Shyanne King	UW M3D	GSR
03/2023 – 02/2026	Serena del Blanco	UW MCB	Member
02/2025 – 02/2026	Samantha Hardy	UW MCB	GSR

(c) TABLE: Advising/Mentoring

Dates	Name	Position	Frequency of Interactions
04/2010 – 08/2018	Brian Oliver, PhD	Postdoctoral Fellow	daily
07/2012 – 10/2018	Vladimir Vigdorovich, PhD	Postdoctoral Fellow	daily
05/2015 – present	Nicholas Dambrauskas	Research Technician	daily
03/2015 – 09/2017	Sara Carbonetti	Research Technician	daily
03/2016 – 09/2018	Emilee Bergle (UW)	Undergraduate Student	weekly
10/2017 – 12/2019	Ramya Chandrasekaran, PhD	Postdoctoral Fellow	daily
05/2017 – 09/2017	Rowan Callahan (Cornell University)	Undergraduate Student	weekly
03/2017 – 01/2022	Olesya Trakhimets	Research Technician	daily
05/2018 – 09/2018	Katy Ni (UW)	Undergraduate Student	weekly
11/2018 – present	Vladimir Vigdorovich, PhD	Research Scientist	daily
01/2019 – 09/2022	Suzannah Beeman (UW)	Undergraduate Student	daily
03/2019 – present	Ganesh Visweswaran, PhD	Research Scientist	daily
06/2019 – 08/2021	Daniela Andrade, PhD	Postdoctoral Fellow	daily
10/2022 – 06/2024	Suzannah Beeman (MS, UW Bioengineering)	Graduate Student	daily
07/2022 – present	Shawn Herring (Ph.D., UW Pathobiology)	Graduate Student	daily

(d) TABLE: Curriculum and Training Program Development

Dates	Role	Responsibility % if shared
None		

(e) TABLE: Other Activities

None	
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11. EDITORIAL RESPONSIBILITIES

TABLE: Editorial Boards

11/2009 – 12/2019	Faculty Member, Faculty of Medicine 1000
11/2011 – 05/2018	Editorial Board Member, AIDS Research and Human Retroviruses
03/2015 – present	Review Editor, Frontiers in Immunology

TABLE: Ad hoc Reviewing Responsibilities

01/2012 – present	Miscellaneous Journal peer review activities – Nature, Science, Immunity, Science Translational Medicine, Cell Host & Microbe, Journal of Virology, Virology, Journal of Immunological Methods, PLoS One, PLoS Pathogens, AIDS research and Human Retroviruses, Science Reports, Nature Communications, NPJ Vaccines, and several others.
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12. SPECIAL RESPONSIBILITIES AND SERVICE

TABLE: International

06/2015 – 07/2018	Michael Smith Foundation research grant reviewer (Canada)
05/2015 – present	Medical Research Council grant reviewer (UK)

TABLE: National

04/2015	NIH special emphasis study section - Glycomics in HIV-1 vaccine research
07/2018	NIH Study Section – HIV Vaccine Research and Design (HIVRAD)
07/2018	NIH Study section – AIDS Immunity and Pathogenesis ad hoc member
06/2019	NIH special emphasis study section – Basic Research in Fc effector function
06/2020	Chair, IPCAVD special emphasis study section
07/2020 – 08/15/2024	Member, HIV Immunopathogenesis & Vaccine Development study section, NIH/NIAID
06/2021	Chair, IPCAVD Special Emphasis Panel, NIH/NIAID/DIADS
11/2024	Ad hoc member, NIH/NIAID Special Emphasis Panel – HIV P01
03/2025	Chair, Special Emphasis Panel on Viral Infections, NIH/NIAID
02/2026	Chair, Special Emphasis Panel on Drugs and Therapeutics, NIH/NIAID
04/2026	Ad hoc member, Special Emphasis Panel, NIH/NIAID

TABLE: Regional

06/2015 – 10/2018	Organizer, Global Health Seminar Series, CIDR/Seattle BioMed
02/2016 – present	Center for AIDS Research Immunology Core associate director, UW/Fred Hutch CFAR, funded by NIH
03/2016 – 10/2018	Translational and Intellectual Property Committee, co-Chair, CIDR
06/2017 – 03/2018	Co-organizer, Keck Family Foundation gift request (successful, \$1M), CIDR

07/2017 – 12/2017	Co-organizer, Murdock Family Foundation capital request for single cell instrumentation (successful, \$750K), CIDR
09/2017 – 01/2018	Organizer, Norquist Foundation capital request (successful, \$400K), CIDR
11/2017 – 02/2026	Co-director, Single Cell Systems Biology Core Facility, SCRI/CGIDR
11/2024 – 02/2026	Center Leadership Team, CGIDR, SCRI, shared resources management

TABLE: University of Washington

None	
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13. CLINICAL ACTIVITIES

None

14. RESEARCH FUNDING

Active Funding

R61 AI161811(Khandhar) 04/09/2021-03/31/2026
HDT Bio / NIAID annual direct costs: \$185,000

Engineering the immune response of a self-replicating and adjuvanting RNA HIV-1 vaccine

The major goal of this project is to evaluate the potential of self-replicating RNA vaccines to elicit antibodies against HIV-1.

Role: Sub-award PI

Effort: 5% FTE

5U01AI165455-03 (Stuart) 02/09/2022-01/31/2027
NIH/NIAID annual direct costs: \$123,925 (to SCRI)

Collective Responses to Malaria Vaccination

The focus of this project is to identify the immunological characteristics of protective responses against malaria infection. The Sather lab will focus on assessing B cell responses using serological assays, next-generation sequencing, and monoclonal antibodies.

Role: Co-PI

Effort: 5% FTE

2U19AI128914 - 07 (Stuart) 03/01/2022-02/28/2027
NIH/NIAID annual direct costs: \$38,515 (to SCRI)

Immune responses to malaria, HIV and SARS-CoV-2 infection and immunization

The major goal of this project is to define immune responses to diseases of high global health importance, to serve as prototypes for the development of interventions.

Role: Co-I

Effort: 4% FTE

1 R01 DE 032663-01 (Sodora/Sather)
NIH/NIDCR

01/15/2023-12/31/2027
annual total direct costs: \$146,909

Influence of viral and immune interventions on early events following oral SIV infection The major goal of this project is to identify the effect of immunomodulatory interventions at the oral mucosa during SIV infection on the initiation of antiviral immune responses, with the goal of identifying new pathways to target with treatment or vaccination.

Role: Co-PI
Effort: 15% FTE

R01 AI 183678 (Swearingen/Sather)
Institute for Systems Biology / NIAID

07/05/2024-6/30/2029
annual total direct costs: \$198,111

Defining the immunogenic landscape of early mosquito-stage *P. falciparum* to accelerate malaria transmission-blocking vaccine discovery

The goal of this project is the discovery of novel vaccine targets in the sexual stage parasite forms of *Plasmodium falciparum*.

Role: Co-PI
Effort: 15% FTE

R01 AI181266-01(Sather/Kappe/Julien)
NIAID

11/01/2024-10/31/2029
annual total direct costs: \$273,241

A universal transmission blocking vaccine for Malaria

The goal of this project is the development of a universal transmission blocking vaccine against malaria based on the conserved fusogens HAP2 and HAP2P.

Role: Co-PI (contact)
Effort: 10% FTE

Past Funding

R33 AI089405-05 (Sather)
NIH/NIAID

08/01/2010-07/31/2016
annual direct costs: \$299,998

Novel HIV 1 Envelope immunogens derived from broadly neutralizing plasmas

The goal of this proposal is the development of novel HIV-1 Envelope immunogens derived from the circulating isolates of individuals that developed broadly neutralizing antibodies during the course of natural infection.

Role: PI
Effort: 50% FTE

5P30 AI 027757-28 (Baeten)
NIH/NIAID/UW

06/01/2013-05/31/2018
annual direct costs: \$21,182

Core I: Immunology Core

The Immunology Core serves the needs of the CFAR community for immune-based assays and provides training and assays on a fee-for-service basis to investigators who are CFAR affiliated at the University of Washington and elsewhere. The Humoral Immunity Sub-Core develops and standardizes novel antibody and neutralization assays for HIV-1, HIV-2 and SIV. The Cellular Immunity develops, standardizes, and trains individuals to perform immunophenotyping and functional flow cytometry panels, T cell and NK cell functional assays.

Role: Immunology Sub-core Director

Effort: 0.05% FTE

R01 AI117234-01A1 (Sather)
NIH/NIAID

04/01/2016-09/31/2021 (NCE)
annual direct costs: \$215,305

Infection-blocking antibody targets for malaria

The goal of this project is the development of novel antibody-inducing vaccines targeting the pre-erythrocytic stage of malaria. We will develop new vaccine candidates and test their protective capacity using cutting edge monoclonal antibody technologies to dissect the mechanisms of protection. If successful, our studies will identify new vaccine candidates to bolster the preclinical vaccine pipeline.

Role: Co-PI

Effort: 15% FTE

R01 DE026336 (Sather)
NIH/NIDCR

07/01/2016-05/31/2022 (NCE)
annual direct costs: \$181,369

Harnessing oral mucosa vaccination to drive protective HIV antibody responses

The goal of this grant is to develop a vaccination strategy to deliver recombinant HIV-1 trimeric Envelope (Env) protein vaccines directly to the oral mucosa (buccal mucosa of the cheek) by intra-epithelial (IEp) vaccination. These experiments evaluate iEp vaccination in the absence of any systemic IM vaccine component and utilize adjuvants to assist in forming a depot of Env protein to the oral mucosa.

Role: Co-PI

Effort: 20 % FTE

OPP1185157 (Sather)
Bill and Melinda Gates Foundation/LIMR

12/10/2017-06/09/2018
annual direct costs: \$55,000

Production and anti-poliovirus activity of human monoclonal secretory IgA

Produce and QC two soluble secretory (sigA) molecules for study in small animal models.

Role: PI (Consortium PI)

Effort: 0.0% FTE

Medical Research Award

07/1/2018-06/30/2020

W.F. Keck Foundation

annual direct costs: \$254,250

Using single cell systems biology to evaluate the role of prior infection in vaccine-induced protection

The goal of this project is to elucidate why several promising experimental vaccines have failed to achieve efficacy in the field after they demonstrated high levels of protection in human clinical trials involving pathogen-naïve individuals, despite inducing similar immune responses. We will use a suite of single cell biology approaches to elucidate how changes in the induction of humoral immunity affect vaccine efficacy.

Role: Co-PI

Effort: Unknown

R01 AI134956 (Kappe)

07/09/2018-06/30/2022

NIH/NIAID

annual direct costs: \$54,366

Molecular Determinants of Sporozoite/Host Cell Interactions

The goal is to build on our findings to fully map and delineate the functional interactions of the P52/P36 complex and its cognate host receptors, as well as the TRAP complex and its host receptors, to gain a comprehensive understanding of initial host infection by the sporozoite stage.

Role: Co-Investigator

Effort: 5% FTE

2P30 AI027757-31 (King)

10/01/2018-05/31/2023

NIH/NIAID/UW

annual direct costs: \$28,106

University of Washington/Fred Hutch Center for AIDS Research (CFAR) - Core I: Immunology Core

The Immunology Core serves the needs of the CFAR community for immune-based assays and provides training and assays on a fee-for-service basis to investigators who are CFAR affiliated at the University of Washington and elsewhere. The Humoral Immunity Sub-Core develops and standardizes novel antibody and neutralization assays for HIV-1, HIV-2 and SIV.

Role: Immunology Sub-core Director

Effort: 5% FTE

R01 AI137234 (Sather)

01/01/2019-12/31/2023

NIH/NIAID

annual direct costs: \$257,068

Development of a pre-erythrocytic *P. vivax* vaccine to prevent clinical relapse

The goal of this project is the development of a vaccine targeting the pre-erythrocytic stages of malaria infection to prevent relapse infection by Plasmodium vivax. Because 80-90% of active blood stage *P. vivax* infections come from relapse, and not primary infection, a vaccine against relapse infection would significantly impact transmission worldwide.

Role: PI

Effort: 16% FTE

R01 AI140951 (Sather)

02/20/2019-01/31/2024

NIH/NIAID

annual direct costs: \$612,114

Kinetics, evolution, and effector function of Fc repertoires during vaccination with native-like Env trimers

The focus of this project is to precisely define the kinetics, evolution, and effector function of vaccine-elicited antibodies against HIV-1, with an emphasis on understanding Fc-mediated activity. We hypothesize that the immunogen, route of administration, adjuvant, and timing all have quantifiable effects on how Fc repertoires are induced, endure, and evolve.

Role: PI

Effort: 20% FTE

5P30AI027757-36 (Celum)

06/01/2023-05/31/2028

University of Washington / NIAID

annual direct costs: \$32,115

University of Washington/Fred Hutch Center for AIDS Research (CFAR) - Humoral Immunity Sub-core

The Humoral Immunity Sub-Core develops and standardizes novel antibody and neutralization assays for HIV-1, HIV-2, SIV, and serves the needs of the CFAR community on a fee-for-service basis to investigators who are CFAR affiliated at the University of Washington and elsewhere.

Role: Core Director

Effort: 5% FTE

15. BIBLIOGRAPHY

(a) Peer-Reviewed Manuscripts

1. Pfent C, Pobursky KJ, **Sather DN**, Golenberg EM. 2005. Characterization of SpAPETALA3 and SpPISTILLATA, B class floral identity genes in *Spinacia oleracea*, and their relationship to floral dimorphism. *Development Genes and Evolution*. 2005 Mar;215(3):132-42. Epub 2005 Jan 20. PMID: 15660251.
2. **Sather DN**, York A, Pobursky KJ, Golenberg EM. Sequence evolution and sex-specific expression patterns of the C class floral identity gene, SpAGAMOUS, in dioecious *Spinacia oleracea* L. *Planta*. 2005 Oct;222(2):284-292. Epub 2005 Jun 7. PMID: 15940462.
3. **Sather, DN**, Armann J, Ching LK, Mavrantoni A, Sellhorn G, Caldwell Z, Yu X, Wood B, Self S, Kalams S, Stamatatos L. Factors associated with the development of cross-reactive neutralizing antibodies during HIV-1 infection. *J. Virol*. 2009 Jan;83(2):757-769. Epub 2008 Nov 5. PMID: 18987148; PMCID: PMC2612355.
4. **Sather DN**, Golenberg EM. Duplication of AP1 within the *Spinacia oleracea* L. AP1/FUL clade is followed by rapid amino acid and regulatory evolution. *Planta*. 2009 Feb;229(3):507-21. Epub 2008 Nov 13. PMID: 19005675.

5. Golenberg EM, **Sather DN**, Hancock LC, Buckley KJ, Villafranco NM, and Bisaro DM. Development of a gene silencing DNA vector derived from a broad host range gemini virus. *Plant Methods*. 2009 Jul 2;5:9. PMID: 19573239; PMCID: PMC2713212.
6. Blish CA, **Sather DN**, Sellhorn G, Stamatatos L, Sun Y, Srivistava I, Barnett SW, Cleveland B, Overbaugh J, and Hu S. 2009. Comparative Immunogenicity of subtype a human immunodeficiency Virus Type 1 envelope exhibiting differential exposure of conserved neutralization epitopes. *J Virol*. 2010 Mar;84(5):2573-84. Epub 2009 Dec 16. PMID: 20015987; PMCID: PMC2820908.
7. **Sather, DN**, Jovanovic, M, Golenberg, EM. Functional analysis of B and C class floral organ genes in spinach demonstrates their role in sexual dimorphism. *BMC Plant Biol*. 2010 Mar 12;10:46. PMID: 20226063; PMCID: PMC2923521.
8. **Sather DN**, and Stamatatos, L. Epitope specificities of cross-reactive NAbS in HIV+ human plasma. *Vaccine*. 2010 May 26;28 Suppl 2:B8-12. PMID: 20510750; PMCID: PMC2879344.
9. Correia BE, Ban YE, Holmes MA, Xu H, Ellingson K, Kraft Z, Carrico C, Boni E, **Sather DN**, Zenobia C, Burke KY, Bradley-Hewitt T, Bruhn-Johannsen JF, Kalyuzhniy O, Baker D, Strong RK, Stamatatos L, Schief WR. Computational design of epitope scaffolds allows induction of antibodies specific for a poorly immunogenic HIV vaccine epitope. *Structure*. 2010 Sep 8;18(9):1116-26. PMID: 20826338.
10. Mikell I, **Sather DN**, Kalams SA, Altfeld M, Alter G, Stamatatos L. Characteristics of the earliest cross-neutralizing antibody response to HIV-1. *PLoS Pathogens*. 2011 Jan 13;7(1):e1001251. PMID: 21249232; PMCID: PMC3020924.
11. Burgers WA, Manrique A, Masopust D, McKinnon LR, Reynolds MR, Rolland M, Blish C, Chege GK, Curran R, Fischer W, Herrera C, **Sather, DN**. Measurements of immune responses for establishing correlates of vaccine protection against HIV. *AIDS Research and Human Retroviruses*. 2012 Jul;28(7):641-8. Epub 2011 Sep 23. PMID: 21861777; PMCID: PMC3380381.
12. Klein F, Gaebler C, Mouquet H, **Sather DN**, Lehmann C, Scheid JF et al. Broad neutralization by a combination of antibodies recognizing the CD4 binding site and a new conformational epitope on the HIV-1 envelope protein. *J Exp Med*. 2012 Jul 30;209(8):1469-79. Epub 2012 Jul 23. PMID: 22826297; PMCID: PMC3409500.
13. **Sather, DN**, Carbonetti S, Kehayia J, Kraft Z, Mikell I, Scheid J, Klein, F., Stamatatos, L. Broadly neutralizing antibodies developed by an HIV+ elite neutralizer exact replication fitness cost to the contemporaneous virus. *J Virol*. 2012 Dec;86(23):12676-85. Epub 2012 Sep 12. PMID: 22973035; PMCID: PMC3497623.
14. Hoot S, McGuire AT, Cohen KW, Strong RK, Hangartner L, Klein F, Diskin, R., Scheid, J.F., **Sather DN**., Burton D.R., Stamatatos L. Recombinant HIV envelope proteins fail to engage germline versions of Anti-CD4bs bNAbs. *PLoS Pathogens*. 2013 Jan;9(1):e1003106. Epub 2013 Jan 3. PMID: 23300456; PMCID: PMC3536657.

15. Diskin R, **Sather DN**, Klein F, Horwitz JA, Halper-Stromberg A, Marcovecchio PM, Lee T, West AP, Gao H, Seaman MS, Stamatatos L, Nussenzweig MC, Bjorkman PJ. Restricting HIV-1 pathways for escape using rationally designed anti-HIV antibodies. *J Exp Med*. 2013 Jun 3;210(6):1235-49. Epub 2013 May 27. PMID: 23712429; PMCID: PMC3674693.
16. Carbonetti S, Oliver BG, Glenn J, Stamatatos L, **Sather DN**. Soluble HIV-1 Envelope immunogens derived from an elite neutralizer elicit cross-reactive V1V2 antibodies and low potency neutralizing antibodies. *PLoS One*. 2014 Jan 23;9(1):e86905. PMID: 24466285; PMCID: PMC3900663.
17. **Sather DN**, Carbonetti S, Malherbe DC, Pissani F, Stuart AB, Hessell AJ, Mikell I, Kalams SA, Haigwood NL, and Stamatatos L. Emergence of broadly neutralizing antibodies and viral co-evolution in two subjects during the early stages of infection with the human immunodeficiency virus type 1. *J Virol*. 2014 Nov;88(22):12968-81. Epub 2014 Aug 13. PMID: 25122781; PMCID: PMC4249098.
18. Harupa A, Sack B, Lakshmanan V, Arang N, Douglass A, Oliver BG, Stuart AB, **Sather DN**, Lindner S, Hybiske K, Torii M, and Kappe S. SSP3 is a novel Plasmodium sporozoite surface protein with a role in gliding motility. *Infection and Immunity*. 2014 Nov;82(11):4643-53. Epub 2014 Aug 25. PMID: 25156733; PMCID: PMC4249349.
19. Malherbe D, Pissani F, **Sather DN**, Guo B, Pandey S, Sutton W, Stuart A, Robins H, Park B, Krebs S, Schuman J, Kalams S, Hessell AJ, and Haigwood NL. Envelope Variants Circulating as Initial Neutralization Breadth Developed in Two HIV-Infected Subjects Stimulate Multiclade Neutralizing Antibodies in Rabbits. *J Virol*. 2014 Nov;88(22):12949-67. Epub 2014 Sep 10. PMID: 25210191; PMCID: PMC4249069.
20. Keitany S, Sack B, Smithers H, Chen L, Jang I, Sebastian L, Gupta M, **Sather DN**, Vignali M, Kappe S, Vaughan A, Wang R. Immunization of mice with live-attenuated late liver stage-arresting malaria parasites generates protective antibody responses to pre-erythrocytic stages. *Infection and Immunity*. 2014 Dec;82(12):5143-53. Epub 2014 Sep 29. PMID: 25267837; PMCID: PMC4249261.
21. Voronin Y, **Sather DN**. Opening the envelope. *IAVI report*. 2015 19(1):16-9. PMID: 26043500.
22. Sampath S, Brazier AJ, Avril M, Bernabeu M, Vidgorovich V, Mascarenhas A, Gomes E, **Sather DN**, Esmon CT, Smith JD. *Plasmodium falciparum* adhesion domains linked to severe malaria differ in blockade of endothelial protein C receptor. *Cellular Microbiology*. 2015 Dec;17(12):1868-82. Epub 2015 Jul 16. PMID: 26118955; PMCID: PMC4661071.
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24. Hessell AJ, Malherbe DC, Pissani F, McBurney S, Krebs SJ, Gomes M, Pandey S, Sutton WF, Burwitz BJ, Gray M, Robins H, Park BS, Sacha JB, LaBranche CC, Fuller DH, Montefiori DC, Stamatatos L, **Sather DN**, Haigwood NL. Achieving potent autologous neutralizing antibody

- responses against Tier 2 HIV-1 viruses by strategic selection of envelope immunogens. *J Immunol*. 2016 Apr;196(7):3064-3078. Epub 2016 Mar 4. PMID: 26944928. PMCID: PMC4797725.
25. *Vigdorovich V, Oliver BG, Carbonetti S, Dambrauskas N, Lange MD, Yacoob C, Leahy W, Callahan J, Stamatatos L, **Sather DN**. Repertoire comparison of the B-cell receptor-encoding loci in humans and rhesus macaques by next-generation sequencing. *Clin Transl Immunology*. 2016 Jul 22;5(7):e93. Doi:10.1038/cti.2016.42. PMID: 27525066; PMCID: PMC4973324.
 26. Nicholas KJ, Flaherty DK, Smith RM, **Sather DN**, Kalams SA. Chronic HIV-1 infection impairs superantigen-induced activation of peripheral CD4⁺CXCR5⁺PD-1⁺cells, with relative preservation of recall antigen-specific responses. *J Acquir Immune Defic Syndr*. 2016 Aug 5. Epub ahead of print. PMID: 27509243; PMCID: PMC5140753.
 27. Yacoob C, Pancera M, Vigdorovich V, Oliver BG, Glenn JA, **Sather DN**, McGuire AT, Stamatatos L. Differences in allelic frequency and CDRH3 region limit the engagement of HIV Env immunogens by putative VRC01 neutralizing antibody precursors. *Cell Rep*. 2016 Nov 1;17(6):1560-1570. PMID: 27806295; PMCID: PMC5207042.
 28. Keitany GJ, Kim KS, Krishnamurty AT, Hondowicz BD, Hahn WO, Dambrauskas N, **Sather DN**, Vaughan AM, Kappe SH, Pepper M. Blood stage malaria disrupts humoral immunity to the pre-erythrocytic stage circumsporozoite protein. *Cell Rep*. 2016 Dec 20;17(12):3193-3205. PMID: 28009289; PMCID: PMC5476299.
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 30. Sack B, Kappe SH, **Sather DN**. Towards functional antibody-based vaccines to prevent pre-erythrocytic malaria infection. *Expert Rev Vaccines*. 2017 May;16(5):403-414. PMID: 28277097.
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(b) MedEdPORTAL or other Peer-Reviewed Curricula

None

(c) Book Chapters

None

(d) Published Books, Video, Software

None

(e) Other Publications

None

(f) Submitted Manuscripts

1.
Oral mucosal vaccination with HIV-1 Env SOSIP elicits potent systemic functional immunity.
2. A novel site of vulnerability on Circumsporozoite protein provides protection from Plasmodium vivax malaria. – revision is under review
3. Mucosal vaccination with HIV-1 Env elicits lymphoid CD4 T cell responses with distinct phenotypes.
4. Failure of B cells to undergo somatic mutation underlies lack of antiviral function, despite germinal center reactions

(g) Abstracts

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9. **Sather DN**, Mikell I, and Stamatatos L. “The earliest cross-neutralizing responses”. *The Search for Broadly Protective Anti-HIV Antibodies*. NIH/NIAID, Bethesda MD, 06/2010.
10. **Sather DN**, Mikell I, and Stamatatos L. “Characteristics of the earliest cross-neutralizing antibody responses in acute infection” *US-Japan AIDS Panel*, Awaji Island, Japan, 9/2010.
11. **Sather DN**, Kraft Z, Kehayia J, and Stamatatos L. “Escape from autologous anti-CD4-BS NAb during natural HIV-1 infection”. *Keystone Symposium*, Whistler British Columbia, CA. 3/2011.
12. **Sather DN**, Carbonetti S, Kehayia J, Kraft Z, Mikell I, Scheid J, Klein F, Nussenzweig MN, Stamatatos L. “Escape from broadly neutralizing antibodies impacts the fitness of the circulating HIV-1 virus in an elite neutralizer”. *Keystone Symposium*, Keystone CO, USA. 4/2012

13. Oliver BG, Carbonetti S, Glenn J, Stamatatos L, **Sather DN**. “HIV-1 gp140 Env immunogens derived from an elite neutralizer elicit conformational V1V2 antibodies and broad, but low potency bNAbs responses”. Keystone Symposium, Keystone CO, USA. 3/2013.
14. **Sather DN**, Oliver BG, Vigdorovich V, Carbonetti S, Lange MD, Lathia K, Saxena A, and Stamatatos L. “B cell receptors containing key structural VRC01-like germline amino acid residues are present in the circulating IgM+ B cell repertoire in outbred rhesus macaques”. Keystone Symposium, Banff Alberta Canada. 3/2014.
15. Vigdorovich VV, **Sather DN**. “Novel Plasmodium antigens targeting the pre-erythrocytic stages of infection”. Gordon Conference, Girona, Spain. 6/2015
16. Sack B, Kappe SHI, **Sather DN**. “Non-CSP antibodies mediate anti-parasitic activity against pre-erythrocytic malaria infection”. Malaria Vaccines for the World, Leiden, Netherlands. 5/2017
17. Vigdorovich V, **Sather DN**. “A pipeline for the development of novel malaria vaccine antigens.” Malaria Vaccines for the World, Leiden, Netherlands. 5/2017
18. Sack B, Kappe SHI, **Sather DN**. “A potential role for antibodies against liver stage plasmodium infection”. Gordon Conference on Malaria, Les Diableries, Switzerland. 7/2017
19. Fisher B, Sodora DL, **Sather DN**. “Oral vaccination with native-like HIV-1 Env trimers elicits systemic immunity and cross-clade reactive anti-V1V2 antibodies”. Keystone Symposium, Banff, Canada. 1/2019.
20. Fisher B, **Sather DN**, Sodora DL. “Oral iEP vaccination with HIV-1 Env trimers”. NHP AIDS 2019, Portland Oregon. 9/2019.
21. Smedley JV, Sodora DL, **Sather DN**. “Use and optimization of Near Infrared (NIR) imaging to guide lymph node collection in macaque models”. NHP AIDS 2021, Portland Oregon. 9/2021

(h) Patents

1. Kappe SHI, Kaushansky A, **Sather DN**. Compositions and Methods for Modulating Eph Receptor Binding and Signaling. WO 2106/123575. File date: 2016-01-29.
2. Kappe SHI, Zanghi G, Flannery E, **Sather DN**. Plasmodium vivax male gamete fusion protein PvHAP2 and the putative proliferating-cell nucleolar antigen P120 as relapse biomarkers. 17/810,982. File date: 2022-7-6.
3. **Sather DN**, Visweswaran GR, Vigdorovich V, Kappe SHI. Multi-stage vaccine for malaria. 63/469,320. File date 2023-5-26

16. TALKS AND PRESENTATIONS

(a) International

10/28/2014 – 10/31/2014	Presenter. “Development of broadly neutralizing anti-HIV-1 antibodies during natural infection through early epitope acquisition and subsequent antibody maturation,” Research 4 Prevention, Cape Town, South Africa
02/08/2015 – 02/13/2015	Presenter. “Repertoire analysis of the B cell receptor-encoding loci in humans and rhesus macaques by next generation sequencing,” Keystone Symposium, Banff, Alberta, Canada.
12/07/2016 – 12/09/2016	Presenter. “Novel antibody-inducing vaccine targets against pre-erythrocytic malaria,” Joint International Tropical Medicine Meeting, Bangkok, Thailand.
06/11/2017 – 06/14/2017	Presenter. “Exploring the potential of vaccine-elicited antibodies against the pre-erythrocytic stages of <i>P. vivax</i> malaria to bolster eradication efforts,” International Conference on <i>P. vivax</i> Research, Manaus, Brazil.
12/06/2017 – 12/08/2017	Presenter. “Toward rational vaccine design for pre-erythrocytic malaria vaccines,” Joint International Tropical Medicine Meeting, Bangkok, Thailand.
08/19/2018 – 08/24/2018	Presenter. “Characterization of a new host receptor interaction for PfTRAP identifies a new site of vulnerability for vaccine design,” ICOPA2018, Daegu, South Korea.
12/12/2018 – 12/14/2018	Presenter. “Toward an anti-relapse vaccine to drive eradication efforts against <i>Plasmodium vivax</i> ,” Joint International Tropical Medicine Meeting, Bangkok, Thailand.
12/07/2022 – 12/09/2022	Presenter. “Advances in pre-erythrocytic vaccines against malaria,” Joint International Tropical Medicine Meeting, Bangkok, Thailand.
03/19/2023	Presenter. “Is a Pan- <i>Plasmodium</i> vaccine an achievable goal?” Convening on Malaria Multi-stage Interventions, Bill and Melinda Gates Foundation, London, England.
12/11/2023 – 12/14/2023	Session chair and Presenter. “Antibodies targeting a novel site of vulnerability on <i>P. vivax</i> CSP reduces hypnozoite formation.” Joint International Tropical Medicine Meeting, Bangkok, Thailand.
12/11/2024 – 12/14/2024	Session co-chair. “Advances in Pre-erythrocytic Prevention”. Joint International Tropical Medicine Meeting, Bangkok, Thailand.
12/07/2025 – 12/09/2025	Session co-chair. “Interventions to prevent malaria infection”. Joint International Tropical Medicine Meeting, Bangkok, Thailand.

(b) National

06/11/2017	Presenter. “Broadly neutralizing antibody responses against HIV-1,” University of California, Davis, Davis, CA.
12/1/2017	Keynote Speaker. “Deciphering host pathogen interactions to inform vaccine design” Wayne State University, Detroit, MI.
01/12/2021	Presenter. “Non-neutralizing anti-CSP antibodies interfere with protective efficacy against malaria”. University of Georgia, Athens, GA.
02/21/2025	Invited Talk. “Features of antibody-mediated protection from malaria infection” University of South Florida, Tampa, FL.

(c) Regional

06/12/2016	Presenter. “Antibody responses induced by Oral vaccination with HIV-1 Env trimers,” Vaccine and Gene Therapy Institute, Beaverton OR.
08/2/2019	Presenter. “Vaccine modalities modulate anti-Env IgG-Fc repertoires and effector responses,” Vaccine and Gene Therapy Institute/OHSU, Beaverton, OR.

(d) Local

05/16/2015	Invited Speaker. “Novel antibody-inducing vaccine targets against pre-erythrocytic malaria,” Fred Hutchinson Cancer Research Center VIDD, Seattle WA
04/24/2019	“Scientific communication: the art of expressing your science”. Pathobiology, University of Washington, Seattle, WA.
02/11/2020	“Untangling humoral immunity against pre-erythrocytic malaria” UW/CERID, Seattle, WA.
09/21/2020	“Decline of neutralizing antibody responses following mild COVID-19 infection” Allen Institute for Immunology, Seattle, WA.
04/14/2021	“Crafting a Successful Vaccine During a Pandemic: An Inside Look at the COVID-19 Vaccines and Seattle Children’s Role.” Science Insights, Seattle Children’s Research Institute, Seattle, WA.

17. OTHER EMPLOYMENT

07/2002 – 09/2002 Summer research fellowship, Wayne State University, College of Science, Detroit, MI

