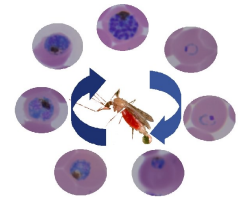


JOHN HOWARD ADAMS

Curriculum Vitae



Center for Global Health & Interdisciplinary Research (GHIDR)
USF Genomics Program, Department of Global, Environmental and Genomic Health Science
College of Public Health, University of South Florida, Tampa, Florida USA



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ORCID <http://orcid.org/0000-0003-3707-7979>
eRA Commons ID: 1888906

Education:

Ph.D., Veterinary Medical Science, University of Illinois (1986)
M.Sc., Veterinary Medical Science, University of Illinois (1982)
B.A., Hendrix College, Conway, Arkansas (1978)

Research and Professional Experience:

Distinguished University Professor (2016 – present), **Distinguished USF Health Professor** (2014 – present), **Professor** (2007 to 2014), Department of Global, Environmental, and Genomics Health Sciences, College of Public Health, University of South Florida, Tampa, Florida and (secondary appointments) Department of Molecular Medicine, USF College of Medicine and Division of Infectious Disease & International Medicine, Department of Internal Medicine, USF College of Medicine.

Professor (2005 to 2007), **Associate Professor** with tenure (1998-2005), **Assistant Professor** (1991-1998), Department of Biological Sciences, University of Notre Dame, Notre Dame Indiana.

Senior Staff Fellow (1989 - 1991), **Staff Fellow** (1987 - 1989), Malaria Section, Laboratory of Parasitic Diseases, National Institute of Allergy and Infectious Diseases, National Institutes of Health, Bethesda, Maryland

Post-doctoral Research Fellow (1986-1987), Department of Parasitology, University of Queensland, St. Lucia, Brisbane, Queensland, Australia

Graduate Assistant (1979-1986), Department of Veterinary Pathobiology, University of Illinois at Urbana-Champaign

Teaching Assistant (1976, 1978), Department of Biology, Hendrix College, Conway, Arkansas

Professional Societies:

- American Society for Microbiology
- American Society of Parasitologists
- American Society of Tropical Medicine and Hygiene

Publication Summary:

Current total peer-reviewed publications, 208; No. of citations: >14,349; h-index: 60; j10-index, 208; citation rate, 800-1000 cites/year (Google Scholar).

Awards:

- 2025 William Trager Award for Basic Parasitology (ASTMH-ACMCIP)
- 2017 Fellow of ASTMH
- 2017 AAAS Fellow (Section on Biological Sciences)
- 2016 USF Distinguished University Professor
- 2014 Distinguished USF Health Professor
- 1997 Burroughs Wellcome Fund New Investigator Award in Molecular Parasitology
- 1986 University of Queensland Postdoctoral Research Fellowship
- 1981 University of Illinois List of Teachers Ranked as Excellent by Their Students
- 1978 Honorable Mention, National Science Foundation Graduate Fellowship

Activities:

- 2018 – now Co-Director, Center for Global Health & Infectious Diseases Research, University of South Florida, Tampa, Florida.
- 2017 – now Director, USF Genomics Program University of South Florida, Tampa, Florida.
- 2006 – now *Trends in Parasitology*, Advisory Board.
- 2020 – 2024 Member, Chair of USF Senate Research Council
- 2009 – 2024 Scientific Advisory Team, PlasmoDB Plasmodium Genomics Resource
- 2017 – 2021 Member, Clinical Research and Field Studies in Infectious Diseases Study Section
- 2020 Co-Chair, ASTMH Basic Science Pre-meeting Course: [The Science and Business of Vaccines Against Tropical Parasitic Diseases in the COVID19 Era](#), November 13, 2020.
- 2009 – 2019 Editor, *Infection and Immunity*
- 2011 – 2017 Mentor, PRIDE – Functional and Applied Genomics of Blood Disorders, NHLBI-Training Program for Junior Faculty at Georgia Health Sciences University
- 2015 Organizing Committee, International Conference on Research for *Plasmodium vivax*, May 19-22, 2015, Bali, Indonesia; June 11-14, Manaus, Brazil.
- 2015 Co-Chair, ASTMH Basic Science Pre-meeting Course: Recent Advances in *In vivo* and *In vitro* Models for Understanding Host-Parasite Interactions, October 25, 2015.
- 2011 – 2015 Member, Chair of USF Senate Research Council
- 2000 – 2014 ASMTM Scientific Program Committee
- 2013 *ad hoc* consultant, NIH Board of Scientific Counselors, site review of LMIV, LMVR & LPD
- 2015 Organizing Committee, Advances in *Plasmodium vivax* Malaria Research & Interdisciplinary Workshops, May 28-30, Barcelona, Spain.
- 2012 – 2013 Visiting Professor, Faculty of Tropical Medicine, Mahidol University, Bangkok, Thailand
- 2008 – 2012 PATH Malaria Vaccine Initiative's Vaccine Science Portfolio Advisory Council.
- 2012 Site review of: Laboratory of Emerging Pathogens, FDA CBER.
- 2010 – 2011 Mentor, SIPID – Functional Genomics of Blood Disorders, NHLBI-Training Program for Junior Faculty at University of Texas at Dallas
- 2009 – 2011 Expert Advisory Group, CRIMALDDI project, Liverpool School of Tropical Medicine
- 2007 – 2010 Pathogenic Eukaryotes Study Section (NIH), Regular Member
- 2009 Member ad hoc, FDA Blood Products Advisory Committee Meeting, November 16, 2009.

- 2009 Chairman, Conference Organizing Committee, *Vivax malaria research III: 2009 and beyond*, Gamboa, Panama, May 24-28, 2009.
- 2008 21st Century World Class Scholar, State of Florida Board of Governors
- 2008 Hendrix College Odyssey Award for Research
- 2000 – 2009 *Infection and Immunity*, Editorial Board member
- 2000 – 2007 ASP R. Barclay McGhee Memorial Lecture Committee, Chair (<http://asp.unl.edu/index.php>)
- 2002 – 2006 Founding President, American Committee of Molecular, Cellular, & Immuno-Parasitology (ACMCIP) of ASTMH (<http://www.astmh.org/subgroup/mp.asp>)
- 2006 ASP Tellers Committee, Chair
- 1998 – 2004 Member / Chair, Scientific Advisory Committee, NIH Malaria Research and Reference Reagent Resource Center
- 2000 – 2004 ASMTH: Young Investigator Award Committee
- 2000 Program Officer, Annual Midwestern Conference of Parasitologists, University of Notre Dame
- 1997 Co-Chair, Planning Meeting for NIAID Malaria Research and Reference Reagent Repository (MR4)
- 1997-1998 Panel Member, USDA National Research Initiative Competitive Grants Program
- Ad hoc reviews: (Journals) *American Journal of Tropical Medicine and Hygiene*, *Clinical & Vaccine Immunology*, *Eukaryotic Cell*, *European Journal of Cell Biology*, *Experimental Parasitology*, *Gene*, *Genomics*, *Infection and Immunity*, *International Journal for Parasitology*, *Journal of Biological Chemistry*, *Journal of Infectious Diseases*, *Journal of Parasitology*, *Journal of Cell Biology*, *Journal of Eukaryote Microbiology*, *Journal of Molecular Biology*, *Malaria Journal*, *Microbes & Infection*, *Molecular and Biochemical Parasitology*, *Molecular Microbiology*, *Nature*, *Nature Communications*, *Nature Methods*, *Nature Structural & Molecular Biology*, *Parasitology Research*, *PLoS ONE*, *PLoS Pathogens*, *PLoS Neglected Tropical Diseases*, *Proceedings of the National Academy of Sciences (USA)*, *Scientific Reports*, *Vaccine*. (Grants) Canada Foundation for Innovation, Human Frontier Science Program; NHMRC (Australia); MRC (UK); NIH (USA) CRFS, IRID, DDR, PTHE, others; USDA National Research Initiative Competitive Grants Program; The Wellcome Trust; Natural Sciences and Engineering Research Council of Canada; Jeffress Memorial Trust.

Research Projects Ongoing or Completed During the Last 3 Years:

Active - research

- | | | |
|--------------------------------------------------------------------------------------------------------------------|-------------------------|------------------------|
| WT-223210/Z/21/Z (Marti, PI; Adams Co-I) | 06/01/23 - 02/28/24 | 5% |
| Wellcome Trust subaward | | \$120,394/ total costs |
| UNRAVELING MECHANISMS OF STAGE CONVERSION IN MALARIA PARASITES | | |
| This project seeks to identify mechanisms sexual commitment in <i>Plasmodium falciparum</i> . | | |
| 2R56AI130171-05A1 (JHA) (Adams, PI) | 06/15/2018 – 06/30/2026 | 15% |
| NIH/NIAID | | \$543,940 TOTAL COSTS |
| DISCOVERING THE ESSENTIAL GENOME OF PLASMODIUM FALCIPARUM | | |
| The objective is to identify and validate genes essential for <i>Plasmodium falciparum</i> gametocyte development. | | |
| R01 AI117017 (Adams, PI; Jiang, Rayner, Otto, Ferdig, Co-I) | 07/12/2021 – 06/30/2026 | 15% |

John H. Adams - Curriculum Vitae

NIH/NIAID \$3,402,299 total /5 YEARS TOTAL COSTS
CHEMOGENOMIC PROFILING OF PLASMODIUM FALCIPARUM DRUG RESPONSES AND RESISTANCE
The objective is to identify and validate gene(s) associated with drug mechanisms of action.

U01 AI155361 (Adams, PI; Cui, Chootong, Popovici, Mao, Ntumngia, Pichyangkul,
Vesely, Vasquez, Waters, Wilder, co-I) 12/01/2020 – 11/30/2026 5%
NIH/NIAID \$4,863,036 total costs

ACCELERATING DISCOVERY OF AN EFFICACIOUS PLASMODIUM VIVAX MULTIVALENT MULTI-STAGE VACCINE.
This project seeks to develop a multi-stage, multivalent vaccine to prevent *P. vivax*.

R01 AI187153 (JHA) (Adams, PI; Marti, Otto, Co-I) 09/01/2024 – 08/31/2029 15%
NIH/NIAID \$3,461,097 TOTAL COSTS

PHENOTYPE-BASED SCREENS TO IDENTIFY GENETIC FACTORS ASSOCIATED WITH GAMETOCYTE DEVELOPMENT IN
PLASMODIUM FALCIPARUM
The objective is to identify and validate genes essential for *Plasmodium falciparum* gametocyte
development

R01 AI183599-01A1 SUBAWARD (HARRISON, PI; DJIMDE ABDOULAYE, CO-I; ADAMS, CO-I)
12/01/2024 – 11/30/2029 2%
NIH/NIAID \$576,594 TOTAL COSTS

GENOMIC SURVEILLANCE FOR ARTEMISININ RESISTANCE IN AFRICA: MOVING BEYOND A CANDIDATE GENE APPROACH
The objective is to evaluate a genomic surveillance approach to identify mutations in functional pathways.

PENDING – RESEARCH

R01 AI179902-01A1 (Adams PI; Kocken, Voorberg, Zeeman, Oberstaller Co-I) -%
NIH/NIAID \$ 3,288,134 total costs

ELUCIDATING ESSENTIAL GENES IN THE LIFE CYCLE OF THE RELAPSING MALARIA PARASITE PLASMODIUM
CYNOMOLGI BY FORWARD GENETICS
This project will apply forward genetic screens to identify genetic factors affecting hypnozoite
development in the NHP relapsing malaria model *P. cynomolgi*.

Active – training

K23 HL150300 (Ho, Thao PI; Adams, mentor) -%
NIH/NHLBI \$195,858 FY total costs

ENTERAL IRON SUPPLEMENTATION AND INTESTINAL HEALTH IN PRETERM INFANTS

This proposal details a 5-year training plan to prepare the physician candidate, Thao (Tina) Ho, for a
career as an independent, translational investigator who will become a leader in the field of iron
deficiency and anemia of prematurity.

Pending – training

none

Completed projects previous 5 years

R01 AI064478 (Adams, PI; Ntumngia, Tolia, Carvalho, Co-I) 08/01/2006 – 03/31/2026 25%
NIH/NIAID \$6,496,541 total costs

Immunological Characterization of the *P. vivax* DBP

This study will optimize a *Plasmodium vivax* Duffy binding protein for clinical vaccine trials.

R01 AI150944 (Yanow, PI; Adams Co-I) 09/01/20 - 02/28/25 5%
NIH/NIAID subaward \$360,320/ total costs

EXPLOITING CROSS-REACTIVE, CONSERVED EPITOPES IN PLASMODIUM VIVAX TO DEVELOP A VACCINE AGAINST FALCIPARUM PLACENTAL MALARIA

This project seeks to develop a conserved target of heterologous immunity as a vaccine candidate.

TC267 (Adams, Roth, Vega-Rodriguez, MPI) 09/01/2019 – 12/10/2024 2%
Tres Cantos Open Lab Foundation 352,939 GBP total costs

ANTIMALARIAL DRUG DISCOVERY TARGETING PRE-ERYTHROCYTIC STAGES OF PLASMODIUM FALCIPARUM

This project offers a major advance in antimalarial drug discovery by targeting pre-erythrocytic stages to block malaria infection.

R01 AI130171 (JHA) (Adams, PI; Otto, Rayner, Co-I) 06/15/2018 – 05/31/2024
NIH/NIAID \$2,775,060 TOTAL COSTS

DISCOVERING THE ESSENTIAL GENOME OF PLASMODIUM FALCIPARUM

The objective is to identify and validate essential genes of *Plasmodium falciparum*.

R21 AI152260 (Lin, MPI; Adams MPI) 11/17/2020 – 10/31/2024 5%
NIH/NIAID \$143,750 total costs

PLASMODIUM OVALE HYPNOZOITE DEVELOPMENT AND RELAPSE: A COORDINATED IN VIVO IN VITRO STUDY

This project will explore genotype-relapse phenotype relationships of *P. ovale* species.

R01 AI137162 (Goldberg/Tolia, PI; Adams, Co-I) 02/01/2017 – 03/31/2024
NIH/NIAID subaward \$ 1,202,134 TOTAL COSTS

STRUCTURAL VACCINOLOGY AND DESIGN OF NOVEL IMMUNOGENS FOR MALARIA VACCINE DEVELOPMENT

The objective is to optimize of *Plasmodium falciparum* CelTOS vaccine.

R21 AI149730 (Adams, PI; Tarning Co-I) 05/22/20 – 04/30/23
NIH/NIAID \$361,288 total costs

EVALUATION OF IVERMECTIN AS AN ANTIMALARIAL THERAPY AGAINST P. FALCIPARUM LIVER STAGE

This project will explore liver metabolites of drug ivermectin in inhibiting the liver stages of *P. falciparum*.

R01 AI117017 (Adams, PI) 04/01/2015 – 07/11/21
NIH/NIAID \$3,451,531 total /5 YEARS TOTAL COSTS

CHEMOGENOMIC PROFILING OF PLASMODIUM FALCIPARUM DRUG RESPONSES AND RESISTANCE

The objective is to identify and validate gene(s) associated with drug mechanisms of action.

W81XWH-18-C-0075 (PI, Milkie; Adams, Co-I) 06/05/18 – 07/31/20
Mainstream Engineering Corporation/US Army \$249,957 total costs

PASSIVELY REGULATED CRYOGENIC (PARC) VIAL SYSTEM FOR VACCINE DISTRIBUTION AND ADMINISTRATION

Evaluating methods for cryopreservation of *Plasmodium* sporozoites.

BAA-NIAID-DAIT-NIHAI2013164 (Fremont, PI; multi-I) 09/01/2014 – 08/31/2019
NIH/NIAID \$1,099,987 total /5 years

B CELL EPITOPE MAPPING OF VIRAL AND PARASITIC ANTIGENS

A contract application that will provide functional characterization of B cell epitopes of *Plasmodium* & Flaviviruses.

USF-PS-001 (Adams J, PI) 03/15/2018-08/31/2018
MVI PATH \$78,392/0.5 YEARS

Determination of IC50 values for antibodies 667,2A10,317 and pfs25(1245) in inhibiting *Plasmodium* sporozoite motility, Traversal and liver stage sporozoites infection

The objective of this project is to evaluate functional efficacy of vaccine antibodies against *Plasmodium falciparum* sporozoite infectivity and LS development.

F32AI112271 (Oberstaller, J - Trainee) 04/01/2015 – 03/31/2018
NIH/NIAID \$168,198/3yrs

POST-TRANSCRIPTIONAL REGULATION IN THE MALARIA PARASITE BLOOD STAGE

The project will study post-transcriptional gene regulatory mechanisms.

John H. Adams - Curriculum Vitae

GRANT No. OPP#1023643 (Adams, PI) 10/26/2010 – 10/31/2017
Bill and Melinda Gates Foundation \$8,580,640 total /5 years
LONG-TERM CONTINUOUS CULTURE OF *PLASMODIUM VIVAX* BLOOD STAGES
The purpose is to develop continuous culture system for blood-stage *P. vivax*.

R01AI094973 (Adams, PI) 04/01/2011 – 02/29/2017
NIH/NIAID \$1,745,388 total /5 years
A LARGE SCALE TRANSPOSON MUTAGENESIS SCREEN OF *PLASMODIUM FALCIPARUM*
The specific aims are to functionally characterize the *Plasmodium falciparum* genome.

R21AI098098 (Adams, PI) 03/20/2012 – 02/28/2016
NIH/NIAID \$356,838 total /3 YEARS
GENETIC SCREEN FOR *P. VIVAX* CQR
The objective is to identify and validate gene(s) associated with CQR in *P. vivax*.

R21AI105328 (Marti, PI) 02/01/2013 – 01/31/2016
NIH/NIAID \$114,273 total
A FORWARD GENETIC SCREEN TO IDENTIFY DETERMINANTS OF MALARIA STAGE CONVERSION
The objective is to identify genetic determinants of gametocytogenesis of *P. falciparum*. /3 years

GRANT No. OPP#1023643 (Kyle, PI) 11/10/2010 – 10/31/2013
Bill and Melinda Gates Foundation \$2,942,389 total /5 years
3D MICROFLUIDIC HUMAN LIVER MODELS FOR MALARIA DRUGS
The purpose is to develop an *in vitro* liver model predictive of *in vivo* outcomes.

P0001-0001020831 (Adams, PI) 08/22/11 – 08/21/12
The Charles Stark Draper Laboratory, Inc. \$41,726 total
DEVELOPMENT OF A LIVER SINUSOID DEVICE FOR STUDYING *PLASMODIUM* EXOERYTHROCYTIC FORMS AND
ANTIMALARIAL THERAPEUTICS
Doctoral fellowship – Steven Maher

PILOT STUDY PROPOSAL (Adams, PI) 01/28/2014 – 05/31/2014
PATH MVI \$40,000 total /3 months
ASSAYS FOR EVALUATING PRE-ERYTHROCYTIC ANTIGENS USING A 3D LIVER MODEL
The purpose is to evaluate an *in vitro* liver model for evaluating anti-sporozoite antibody..

MMV 12/0076 (Adams, PI) 07/01/2012 – 06/30/2013
Medicines for Malaria Venture \$50,000 total
CHEMICAL PROFILING OF *P. FALCIPARUM* GENETIC MUTANTS TO OPTIMIZE DRUG DEVELOPMENT I
The objective is to elucidate drug mechanisms of action and functionally annotate *P. falciparum* genome.

N66001-11-1-4174 (Deschenes, PI) 08/24/11 – 02/23/14
Defense Advanced Research Projects Agency \$310,766 total
COUNTERMEASURES TO COMBAT PROTOZOAN PARASITES
The goal is to characterize a set of kinase and phosphatase targets specific for *T. gondii* and
Plasmodium; yet are conserved across *Apicomplexa* parasites in general; toward the development of
therapeutic agents against the *Apicomplexan* parasites.

RFP No. GBM-11-10-09 (Adams, PI) 01/28/2010 - 09/21/2013
Science Applications International Corp \$ 693,916 total
EVALUATION OF PVDBPRII IMMUNOGENS FOR IMMUNOGENICITY AND PROTECTIVE EFFICACY TOWARD THE
DEVELOPMENT OF AN ANTI-DBP VACCINE AGAINST *PLASMODIUM VIVAX*
A comparative evaluation of different PVRII vaccine designs.

CDDI Conference grant (Adams, PI) 02/01/12 – 01/31/13
USF System Internal Awards Program \$9,996 total
NEW FRONTIERS OF DRUG DISCOVERY: ACADEMIC RESEARCH

Conference will provide doctoral students with the opportunity to present their infectious disease research as it relates to academic drug discovery.

R01 AI064478 supplement (Adams, PI)

08/01/2009 - 07/31/2012

NIH/NIAID

\$274,379 total

IN VITRO ASSAYS P. VIVAX DBP

The specific aims of this proposal are to characterize residues on the *Plasmodium vivax* Duffy binding protein responsible for antigenic character and sensitivity to neutralizing antibody inhibition.

F31 AI83053 (Adams, mentor)

08/01/2009-07/31/2011

NIH/NIAID

\$30,074 YR1 total

FUNCTIONAL CHARACTERIZATION OF A CONSERVED PHOSPHATASE OF PLASMODIUM FALCIPARUM

Organize on campus symposium and retreat for doctoral research program in Drug Discovery in Infectious Diseases.

Patents issued:

US patent No. 11,787,845 B1 (Oct 23, 2023) "Synthetic Plasmodium Antigens, Compositions, and Uses Thereof." Inventors: JH Adams, CL King, NH Tolia, E Chen, ND Salinas, MY George, FB Ntumngia, SJ Barnes. Provided herein are synthetic *P. vivax* antigens, antibodies and pharmaceutical formulations and vaccines thereof.

US Patent No. 11,331,367 B1 (May 17, 2022) "Compounds and Methods for Inhibiting Protozoan Parasites." Inventors: BJ Baker, NG Wilson, JH Adams, MA Knestrick, AE Roth. Described herein are compounds, formulations, and methods for blocking sporozoite invasion and subsequent liver-stage parasite development of a protozoan parasite, such as *Plasmodium falciparum*

US patent No. 11,332,711 (May 17, 2022) "Sporozoite Cryopreservation Compositions and Methods." Inventors: JH Adams, AE Roth. Invention is for new effective methods to cryopreserve *Plasmodium* and other eukaryotic organisms.

US patent No. 10,927,153 B1 (Feb 23, 2021) "Synthetic Plasmodium Antigens, Compositions, and Uses Thereof." Inventors: JH Adams, CL King, NH Tolia, E Chen, ND Salinas, MY George, FB Ntumngia, SJ Barnes. Provided herein are synthetic *P. vivax* antigens, antibodies and pharmaceutical formulations and vaccines thereof.

US Patent No.: 9,120,869 (Sep. 1, 2015). "Synthetic antigen based on the ligand domain of the *Plasmodium vivax* Duffy binding protein". Inventors: JH Adams, FB Ntumngia, JL Schloegel, SJ Barnes, AM McHenry, P Chootong. Invention relates to a newer version of DBP vaccine to prevent *Plasmodium vivax*.

US Patent No.: 8,784,832 (July 22, 2014). "Synthetic antigen based on the ligand domain of the *Plasmodium vivax* Duffy binding protein". Inventors: JH Adams, FB Ntumngia, JL Schloegel, SJ Barnes, AM McHenry, P Chootong. Invention relates to a DBP vaccine to prevent *Plasmodium vivax*.

US Patent 7,932,088 (April 26, 2011). "High Efficiency Transformation of *Plasmodium falciparum* by the Lepidopteran Transposon, *piggyBac*." Inventors: JH Adams, MJ Fraser, Jr., B Balu, DA Shoue. Invention relates to use of *piggyBac* as a tool for genetic manipulation of the *Plasmodium* genome.

US Patent No. 6,120,770 (September 19, 2000). "*Plasmodium* Proteins Useful for Preparing Vaccine Compositions." Inventors: JH Adams, S Kappe and JP Dalton. Invention relates to *Plasmodium* MAEBL as it can be used as a vaccine for humans against malaria.

US Patent No. 5,541,292 (July 30, 1996). “*Plasmodium vivax* and *Plasmodium knowlesi* Duffy receptor.” Inventors: LH Miller, JH Adams, DC Kaslow, and X Fang. Invention relates to the Duffy binding protein of a *Plasmodium* parasite as it can be utilized as a vaccine for humans against malaria.

US Patent No. 5,198,347 (March 30, 1993; expired). “DNA encoding *Plasmodium vivax* and *Plasmodium knowlesi* Duffy receptor.” Inventors: LH Miller, JH Adams, DC Kaslow, and X Fang. Invention relates to the Duffy binding protein of a *Plasmodium* parasite as it can be utilized as a vaccine for humans against malaria.

Patents applications pending:

US Patent No. 11,331,367 (pending) “Compounds and Methods for Inhibiting Protozoan Parasites.” Inventors: BJ Baker, NG Wilson, JH Adams, MA Knestrick, AE Roth.

US Patent No. 201808544 A1 (pending). “Compositions comprising CelTOS immunogens and antibodies and method of use thereof”. Inventors: N Tolia, J Jimah, N Salinas, D Urusova, JH Adams, S Xu, A Roth, H Kumar. The invention structurally defines critical functional epitopes of *Plasmodium* CelTOS.

US Patent Application No.: 2015/0368599 A1 (December 24, 2015). "Design and hot embossing of macro and micro features with high resolution microscopy access". Inventors: S Maher, WM Sadi, AJ Taylor, HS Sun, D Kyle, J Adams. Invention relates to micro-feature devices and methods for fabricating micro-feature devices.

US Patent Application No. 13/237,525 priority date March 29, 2009. “Method and composition using a dual specificity protein tyrosine phosphatase as an antimalarial drug target”. Inventors: JH Adams, B Balu, SP Maher, C Campbell, R Manetsch. This invention relates to the treatment of malaria. Specifically, this invention relates to the discovery of a novel drug target for the treatment of malaria. Application abandoned by USF Division of Patents & Licensing, 2013.

Courses Taught (UND):

Undergraduate Level:

BIOS 201	General Biology
BIOS 40415	Medical & Veterinary Parasitology
BIOS 415L	Parasitology Lab
BIOS 418*	Molecular Genetics
BIOS 494	Directed Readings for Undergraduates
BIOS 499	Undergraduate Research

Graduate Level:

BIOS 510	Experimental Parasitology
BIOS 514	Field Parasitology Laboratory
BIOS 516	Physiological Chemistry of Animal Parasites
BIOS 60530	Immunobiology of Infectious Diseases/ Advanced Immunology
BIOS 580	Graduate Seminars

University committee service

University of South Florida.

Graduate Council, Policy subcommittee (2009-2011); USF Research Council (2011-2015 and 2021-2024), Chair-elect (2013/2014), Chair (2014/2015 and 2022/2023).

College of Public Health.

Appointments for Promotion and Tenure, Research Committee, various faculty search committees

Other roles.

Founder and Director of USF Genomics Program (2016-current), Co-Director of Center for Global Health and Infectious Diseases Research (2017-current)

Faculty Research Training Record

Postdoctorate level.

1. **M. Dennis Prickett**, NIH Postdoctoral fellows, 1991-1993. Undergraduate degree: Mississippi State University. Ph.D., University of Georgia. Research project: genetic analysis of polymorphisms of the *pkdhp* and *pvdhp*. Position after leaving: 3-year MRC Fellowship studying apical organelles of *Theileria* in the laboratory of Dr. Roger Hall, University of York, UK. Subsequent position: Research Scientist, Cancer Center, Milan, Italy.
2. **Naresh Singh**, M.Sc., Ph.D. Postdoctoral Research Assistant, 1/2001-6/2004 (R01 AI33656; UND 45742). Project: expression, purification and structural analysis of recombinant *Plasmodium falciparum* MAEBL ligand domains. Preceding appointment as Postdoctoral Fellow in the Malaria Research Group, International Centre for Genetic Engineering & Biotechnology, New Delhi, India. Current position, Research Associate, University California at San Francisco
3. **Jun Fu**, Ph.D. Postdoctoral Research Assistant (1/2002-3/2005)(R01 AI33656). Epitope mapping of *Plasmodium falciparum* MAEBL ligand domains. Preceding appointment as Research Fellow, Department of Medical Zoology, Nagoya City University Medical School, Japan
4. **Chitra Chauhan**, Postdoctoral Research Assistant, 2007, (R21 AI07088). Design and analysis of the effects of gene disruption in a genetic screen *Plasmodium falciparum*. PhD, 2005, Institute of Genomics and Integrative Biology, New Delhi, India, in Molecular Genetics. Deceased.
5. **Jesse Schloegel**, Postdoctoral Research Assistant, 2009 - 2010, (R01 AI064478). Identification and characterization of epitopes on the *Plasmodium vivax* Duffy binding protein using phage display. PhD, 2008, LaTrobe University, Bundoora, Australia.
6. **Bharath Balu**, Postdoctoral Research Assistant, 2006 - 2011, (R01 AI33656/ R21 AI07088). Design and apply a genetic screen to identify genes important in *Plasmodium falciparum* sporozoite development in the mosquito. PhD, 2006, University of Notre Dame.
7. **Saranya Siribal**, Postdoctoral Research Associate, 2010, (BMGF *in vitro* models project). Development of continuous *in vitro* blood-stage culture of *Plasmodium vivax*. Accepted position as Research Scientist, Mahidol Vivax Research Unit, Faculty of Tropical Medicine, Mahidol University. Accepted position at Western University, Thailand. PhD, 2010, Mahidol University, Bangkok Thailand.
8. **Wanlapa Roobsoong**, Postdoctoral Research Associate, 2010 - 2012, (BMGF *in vitro* models project). Development of continuous *in vitro* blood-stage culture of *Plasmodium vivax*. Accepted position as Research Scientist, Mahidol Vivax Research Unit, Faculty of Tropical Medicine, Mahidol University. PhD, 2010, Mahidol University, Bangkok Thailand.
9. **Rajeev Tyagi**, Postdoctoral Research Associate, 2011 - 2013, (R01 AI094973; BMGF *in vitro* models project). Development of humanized mouse models for experimental malaria research. PhD, 2011, Institut Pasteur, Paris, France.

10. **Anatoli Naumov**, Research Associate, 2009 - 2014, (BMGF *in vitro* models project). Optimizing expression of the *Plasmodium vivax* Duffy binding protein. PhD, 1985, Russian Academy of Sciences, Pushchino, Moscow Region, Russia.
11. **Naresh Singh**, Research Associate, 2008 - 2015, (BMGF *in vitro* models project). Project: Analysis *Plasmodium falciparum* sporozoite development in wild type and genetic mutant parasite clones. Preceding appointment as Postdoctoral Research Associate, University California at San Francisco and University California, Davis. MSc, PhD.
12. **Richard Thomson-Luque**, Research Associate, 2014 – 2017 (BMGF *in vitro* models project). Establishment of long-term continuous *in vitro* culture of blood-stage *P. vivax*. BMSc University of Málaga 2000; MSc, University Autónoma of Barcelona, 2002.
13. **Sandhya Boyapalle**, Research Associate, 2015 – 2017. (R01AI094973 & BMGF *in vitro* models project). DVM, 1997, Acharya Ng Ranga Agricultural University, MSc, 1999, Tuskegee University, PhD, 2005, Iowa State University. Analysis *Plasmodium falciparum* intraerythrocytic development in wild type and genetic mutant parasite clones; evaluation of *P. vivax* liver stages in humanized mouse model.
14. **Jennifer Armistead**, Postdoctoral scholar, 2014 - 2017, (BMGF *in vitro* models project). Selection and characterization of higher transmitting line of *Plasmodium vivax* from NHP models and development of transgenic reporter lines, a collaboration with Tom Wellems LMVR NIAID. PhD 2013, MPH 2008 Johns Hopkins Bloomberg School of Public Health; MSc 2007 Florida Medical Entomology Laboratory, University of Florida; BSc 2002, George Mason University.
15. **Alison Roth**, Research Associate, 2018. Evaluation of antimalarial liver stage therapies. PhD 2018, University of South Florida College of Public Health; MPH 2013, USF COPH; BSc, Michigan State University.
16. **Sai Lata De**, Postdoctoral scholar, 2017 - 2023, (R01 AI064478). Pre-clinical studies to enhance immunogenicity of an engineered *Plasmodium vivax* Duffy binding protein vaccine. PhD 2016, Institute for Glycomics, Griffith University; MSc 2010, Sri Ramachandra Medical College and Research Centre (SRMC); BSc 2008, Bangalore University.
17. **Prem Prakash**, PhD, Postdoctoral scholar, 2022 – 2024. Scientist, Division of Molecular Microbiology and Immunology, CSIR-Central Drug Research Institute-Lucknow, Jankipuram Extension, Sector-10, Sitapur Road, Lucknow, India 226031.
18. **Francis Ntumngia**, Research Assistant Professor, 2013 – present (R21 AI107455-(PI); R01 AI064478); Postdoctoral Research Assistant/Associate, 2006 – 2013, (R01 AI064478; SAIC contract). Immunochemical characterization of conserved neutralizing epitopes on the *Plasmodium vivax* Duffy binding protein. PhD, 2006, Institut für Tropenmedizin der Universität Tübingen, Germany.
19. **Shulin Xu**, Research Associate, 2012 - present, (R21AI098098; R21AI105328; R01AI094973). Genetic analysis of drug resistance and gene regulation in human malaria parasites. PhD, 1992, Graduate School of Chinese Academy of Agricultural Sciences, Beijing, China
20. **Jenna Oberstaller**, Postdoctoral scholar/Research Associate, 2012 – present, (F32AI112271 Fellowship; previously, R01AI094973 & BMGF *in vitro* models project; R01AI130171, Interdisciplinary Sciences Liaison, USF Genomics Program). Comprehensive analysis of post-transcriptional regulatory mechanisms of *P. falciparum*; bioinformatics analysis of metabolic pathways of *P. vivax* to support development of long-term *in vitro* culture of blood-stage parasites. PhD 2012, University of Georgia, Athens, Georgia.

21. **Camilla Valente Pires**, PhD, Postdoctoral scholar, 2020 – 2024; Research Associate, 2024 - present. Postdoctoral Research Scientist from the Oswaldo Cruz Foundation, René Rachou Research Center, Malaria Laboratory in Belo Horizonte, Brazil. Functional genomics of *Plasmodium falciparum*: evaluation of genes essential for survival of oxidative stress.
22. **Pradeep Subramni**, PhD, Postdoctoral scholar, 2020 – 2025; Research Associate, present.
23. **Surendra Kumar Kolli**, PhD, Postdoctoral scholar, 2020 – 2025; Research Associate, present.

Predoxoratorate level.

1. **Stefan Kappe**, Doctoral student, 1992-1998. Undergraduate degree and Diploma at Universität Bonn. Dissertation title: "Molecular Cloning of *maeb1* of *Plasmodium yoelii yoelii* and *Plasmodium berghei*". Received BBMB Fellowship 93/94 academic year. Awards: LaRue Award for best student oral presentation AMCOP 1997; Outstanding Young Investigator Award, Molecular Parasitology Meeting 1997, Woods Hole; ASTMH Young Investigator Award, 1997 Annual Meeting. Initial position: Recipient of Bernard B. Levine Fellowship, NYU School of Medicine, Laboratory of Dr. Victor Nussenzweig; promoted to Assistant Professor, Department of Pathology, NYU School of Medicine; current position, Associate Member, Seattle Biomedical Research Institute, Seattle, Washington.
2. **Amy R. Noe**, Doctoral student, 1995 to 1999. Undergraduate degree San Diego State University. Dissertation topic: "Partial Characterization of the Protein MAEBL and its Use as a Molecular Marker for Organelle Biogenesis in the Malaria Parasite". Predoxoratorate fellow on NIH Experimental Parasitology and Vector Biology Training Grant, 1/96-6/98. Initial position: Senior Scientist/Project Manager, Allered, San Diego, California; Current position: Senior Biodefense Analyst, SAIC, San Diego, California.
3. **Pascal Michon**, Doctoral student, 1995 to 2001. Undergraduate degree at Université de Poitiers and graduate training at Université Montpellier. Dissertation topic: "Erythrocyte binding proteins of *Plasmodium vivax* and *Plasmodium knowlesi*: molecular and phylogenetic approaches". Initial position: postdoctoral research fellow, The Wellcome Trust Centre for the Epidemiology of Infectious Disease, Department of Zoology, University of Oxford, Laboratory of Prof. Karen P. Day. Current position: Senior Research Fellow, Papua New Guinea Institute for Medical Research, Madang PNG.
4. **Peter Linn Blair**, Doctoral student, 1996 to 2002. Undergraduate degree at Berea College. Dissertation topic: "Molecular cloning and characterization of the *Plasmodium falciparum maeb1*". Predoxoratorate fellow on NIH Experimental Parasitology and Vector Biology Training Grant, 8/96-6/99. Awards: Honorable Mention ASTMH Young Investigator Award, 1999 Annual Meeting; participant, Biology of Parasitism annual course at Marine Biology Laboratory, Woods Hole, MA. Initial position: National Research Council Associate, Naval Medical Research Medical Center, Forest Glen, MD; supervisor Dr. Daniel Carucci. Current position: Assistant Professor, Earlham College.
5. **Anita Vincent**, Doctoral student, 1999 – 2002, Undergraduate degree at St. Joseph's College (India), Graduate degree (M.Sc.) at Loyola University of Chicago. Dissertation topic: Expression and immunogenicity of *Plasmodium* erythrocyte binding proteins. Did not complete degree requirements.
6. **Eunita Ohas**, WHO Predoxoratorate Fellow and Visiting Doctoral student, 1999-2003 from Kenyatta University, Kenya. Dissertation topic: "Immunogenicity of the *Plasmodium falciparum* Erythrocyte Binding Antigen-175". PhD granted posthumously from Kenyatta University, 2003.

7. **Kelley VanBuskirk**, Doctoral student, 1997 to 2004. Undergraduate degree at University of North Carolina-Chapel Hill. Dissertation topic: “Partial immunological and functional characterization of the *Plasmodium vivax* Duffy binding protein”. Awards: Recipient of a University of Notre Dame Luce Fellowship for Outstanding Woman Graduate Student; Honorable mention for best student poster presentation AMCOP 2000 & AMCOP2002. Current position: Post doc, Seattle Biomedical Research Institute
8. **Bharath Balu**, Doctoral student, 2000 – 2005, Undergraduate degree (India). Dissertation topic: “Genetic Analysis of *Plasmodium falciparum* using *piggyBac*-mediated insertional mutagenesis”. Travel award to participate in 2002 Workshop on Transfection of Malaria Parasites, ICGEB, New Dehli, India. Current position: student in Physician Assistant.
9. **Fabián Ernesto Saenz**, Doctoral student, 2002 – 2008, Undergraduate degree (La Pontificia Universidad Católica del Ecuador), recipient of *Fulbright/Western Hemisphere Fellowship* and Coca Cola Award from University of Notre Dame Kellogg Institute. Dissertation topic: “Genetic analysis of the role of MAEBL for *Plasmodium falciparum* sporozoite invasion of the anopheline salivary gland”. Travel award to participate in Workshop on Microarray applications with Malaria Parasites, Bangkok, Thailand. Current position: Assistant Professor, La Pontificia Universidad Católica del Ecuador.
10. **Sarita Mendonca**, Doctoral student, 2003 – 2008, Undergraduate degree (India), Dissertation topic: “Structural analysis of MAEBL ligand domains of *Plasmodium falciparum* sporozoites and partial characterization of its receptor on *Anopheles* salivary glands”.
11. **Patchanee Chootong**, visiting ‘sandwich’ PhD student from Mahidol University (advisor: Prof. Rachanee Udomsangpetch), Department of Pathobiology. Identification and characterization of epitopes of *Plasmodium vivax* Duffy binding protein using immune sera. November, 2006 – August, 2008. (R01 AI064478)
12. **Amy McHenry**, Doctoral student, 2004 - 2009. Undergraduate degree at Union College, Lincoln, Nebraska. Dissertation topic: “Functional characterization of variant neutralizing epitopes on the *Plasmodium vivax* Duffy binding protein”. Travel award to participate in Workshop on in vitro cultivation of *P. vivax*, Mae Sot, Thailand. Awards: Recipient of a University of Notre Dame Schmitt Fellowship; Best student presentation AMCOP 2005.
13. **Chris Campbell**, COPH Doctoral student, 2007 – 2013. Undergraduate degree (B.Sc.) and Graduate degree (M.Sc.) at Andrews University. Berrien Springs, Michigan. Dissertation topic: functional characterization of conserved cell cycle protein tyrosine phosphatase in *P. falciparum*. Recipient of F31AI 83053. Current position: Assistant Professor, Advent Health University.
14. **Jennifer Sedillo**, COPH Doctoral student, 2009 - 2014. Dissertation topic: understanding phosphorylation cascades as a regulator of *Plasmodium falciparum* cell cycle. Recipient of USF Presidential Scholar award. Current: Associate Professor, American Public U. System.
15. **Steven P. Maher**, Draper Lab Fellow /COPH Doctoral candidate, 2009 – 2014. Dissertation topic: development of *in vitro* liver stage models for human malaria parasites that predictive of *in vivo* outcomes. Recipient of Draper Laboratory Fellowship. Research Scientist, University of Georgia CTEGD.
16. **Phaedra Thomas**, COPH Doctoral student, 2008 – 2015. Undergraduate degree (B.S.) Albany State University and Graduate degree (M.S.) at University of South Florida. Dissertation topic: A forward genetic screen identifies factors associated with fever pathogenesis in *Plasmodium falciparum*. Bridge to the Doctorate Fellowship Award, Florida-Georgia Louis Stokes Alliance for Minority Participation.

17. **Miriam T. George**, COPH Doctoral student, 2012 – 2015. Undergraduate degree (B.S.) Vellore Institute of Technology, Vellore, India and Graduate degree (MPH) at University of South Florida. Dissertation topic: Immunological characterization of Duffy binding protein of *Plasmodium vivax*. Project Manager – NSSEFI, Wellcome Trust Research Laboratory, Christian Medical College, Vellore, India.
18. **Alison Roth**, COPH Doctoral student, 2015 – 2018. Undergraduate degree (B.S.) Michigan State University and Graduate degree (MPH) at University of South Florida. Dissertation topic: Assessment of therapeutic targets of sporozoites of *Plasmodium vivax* and *P. falciparum*. Research Microbiologist, Walter Reed Army Institute of Research.
19. **Caroline F. Simmons**, PPIBS Doctoral student, 2017 – 2023. Undergraduate degree (B.S.) University of South Florida. Dissertation topic: Functional genomics of *Plasmodium falciparum* conserved genes of unknown function. Bridge to the Doctorate Fellowship Award, Florida-Georgia Louis Stokes Alliance for Minority Participation. Clinical Scientist, Tampa General Hospital, Tampa, Florida.
20. **Jyotsna Chawla**, PPIBS Doctoral student, 2017 – 2023. Undergraduate degree (B.S.) and graduate degree (M.Sc.) Fergusson College, Savitribai Phule Pune University, Pune, India. Dissertation topic: Large-scale forward genetic screens of *Plasmodium falciparum* piggyBac mutants for aberrant gametocyte phenotypes. Assistant Professor of Medical Microbiology, Department of Foundational Sciences, Dr. Kiran C. Patel College of Osteopathic Medicine, Nova Southeastern University, Clearwater, Florida.
21. **Justin Nicholas**, PPIBS Doctoral student, 2017 – 2023. Undergraduate degree (B.S.) University of North Florida. Dissertation topic: Characterization of *P. vivax* pre-erythrocytic vaccine candidates. Associate Scientist, Iovance Biotherapeutics, Inc., Tampa, Florida.
22. **Olatunbosun O. Aringbangba**, COPH Doctoral student (Genomics), 2022 – present.
23. **Pooya Mahdavi**, COPH Doctoral student (Global Communicable Diseases), 2023 – present.
24. **Chiara Michelli**, PPIBS Doctoral student, 2023 – present.
25. **Mary Avornyo**, PPIBS Doctoral student, 2023 – present.

Visiting scientists:

G. Paul Curley, PhD, Visiting Postdoctoral researcher from Dublin City University, 1992. University College Dublin laboratory. Collaborative research project with John Dalton: Identification and cloning of the *ebf* homologues from rodent malaria parasites.

Takafumi Tsuboi, MD PhD, Visiting Scholar from Ehime University School of Medicine, Japan, 1992-1993. Ehime University School of Medicine. Collaborative research project: molecular epidemiology of *Plasmodium vivax* and genetic analysis of polymorphisms in the *pvdgp* locus. Current position: Director & Professor, Ehime University Proteo-Science Center (PROS).

Bernadette Condon, PhD, Visiting Postdoctoral researcher from Dublin City University, 1995. University College Dublin. Collaborative research project: molecular cloning of the active metalloproteases from rodent malaria parasites.

Tonya Bonilla, visiting graduate student July 24-28, 2006 from University of Florida (advisor: Prof. John Dame), Dept. of Infectious Diseases and Pathology, Gainesville, Florida. Collaboration to develop *piggyBac* tools for malaria research (R21 AI07088).

Capt. Alyson Auliff, Visiting Fulbright Scholar, July 1, 2009 – June 30, 2010. Scientific Officer, Department of Drug Resistance and Diagnostics, Army Malaria Institute, Gallipoli Barracks, Enoggera (Brisbane) QLD. Analysis of *Plasmodium vivax* drug resistance genes by transgene expression in *Plasmodium falciparum*.

Hitoshi Otsuki, MD PhD, Visiting Scholar, April 1, 2011 – March 30, 2012. Scientific Officer, Tottori University, Faculty of Medicine, Department of Medical Zoology, Tottori, Japan. Analysis of *Plasmodium falciparum* gene regulation.

Capt. Alyson Auliff, PhD, Visiting Scientist, 2013. Scientific Officer, Department of Drug Resistance and Diagnostics, Army Malaria Institute, Gallipoli Barracks, Enoggera (Brisbane) QLD. Fulbright Scholar. Application of *piggyBac*-mediated transgenesis in *P. falciparum*.

Flora Kano, PhD, Visiting Scientist, 2013. Research Scientist from the Oswaldo Cruz Foundation, René Rachou Research Center, Malaria Laboratory in Belo Horizonte, Brazil. Collaboration studies with Luzia Carvalho on immunogenicity of PvDBP.

Maja Malmberg, PhD, 2014. Visiting Scientist from Swedish University of Agricultural Sciences. Molecular analysis of drug resistance in malaria parasites. PhD, Karolinska Institutet, Stockholm Sweden. Forward genetic studies *piggyBac*-mutants in *P. falciparum*.

Letícia de Menezes Torres, 2014-2015. Visiting Doctoral student from Doutorado em Ciências da Saúde, Centro de Pesquisas René Rachou/FIOCRUZ - Laboratório de Malária; Luzia Carvalho, major Professor. Collaboration studies with Luzia Carvalho on immunogenicity of *P. vivax* DBP and RBPs.

Kézia K. G. Scopel, PhD, Visiting Scientist 2014-2015. Professora Adjunta from the Instituto de Ciências Biológicas, Universidade Federal de Juiz de Fora. Advanced training in functional genomics of malaria parasites and *in vivo* humanized mouse models for malaria research.

Camilla Valente Pires, PhD, Visiting Scientist, 2015-2016. Postdoctoral Research Scientist from the Oswaldo Cruz Foundation, René Rachou Research Center, Malaria Laboratory in Belo Horizonte, Brazil. Collaboration studies with Luzia Carvalho on immunogenicity of PvDBP, funded jointly by NIH-CNPQ.

Vishal Saxena, PhD, 2015 & 2017, Visiting Scientist from Center for Biotechnology, Molecular Parasitology and System Biology Lab, Department of Biological Sciences, Birla Institute of Technology & Science, Pilani, 333031, Rajasthan, India. Molecular genetic and cell biological studies of *P. vivax*.

Achaporn 'Preaw' Yipsirimetee, PhD candidate, 2023, The Royal Golden Jubilee PhD Programme, The Thailand Research Fund, Faculty of Tropical Medicine, Mahidol University, Bangkok, Thailand.

Lauriane Sollelis, PhD, 2023 & 2025, Visiting Scientist and Research Associate from Institute of Parasitology Zurich (IPZ), University of Zurich, Switzerland. Forward genetic screens to identify genetic regulators of *Plasmodium falciparum* gametocyte development.

Plus 62 University of Notre Dame & University of South Florida undergraduate students, including three who graduated as an ND Outstanding Biology Major and two as Outstanding Honors Program Graduate in Science and four USF Honors students with thesis.

Refereed Publications:

MSc-related publications

1. Todd Jr KS, **Adams JH**, Hoogweg JH. (1978). The muskrat, *Ondatra zibethica*, as a host of *Taenia mustelae* in Illinois. *Journal of Parasitology*. **64**:523.
 2. **Adams JH**, Levine ND, Todd Jr KS. (1981). *Eimeria* and *Sarcocystis* in raccoons in Illinois. *Journal of Protozoology*. **28**:221-222.
 3. Blagburn BL, **Adams JH**, Todd Jr KS. (1982). First asexual generation of *Eimeria vermiformis* Ernst, Chobotar, and Hammond, 1971 in *Mus musculus*. *Journal of Parasitology*. **68**:1178-1180.
 4. **Adams JH**, Todd Jr KS. (1983). Transmission electron microscopy of intracellular sporozoites of *Eimeria vermiformis* (Apicomplexa, Eucoccidiida) in the mouse. *Journal of Protozoology*. **30**:114-118.
 5. Blagburn BL, **Adams JH**, Todd Jr KS, Warner KA. (1983). Prevalence of heartworm in dogs. A survey of southwestern Michigan and northern Indiana. *Modern Veterinary Practice*. **64**:811-814.
 6. **Adams JH**, Todd Jr KS. (1984). Transmission electron microscopy of meront development of *Eimeria vermiformis* Ernst, Chobotar and Hammond, 1971, (Apicomplexa, Eucoccidiorida) in the mouse, *Mus musculus*. *Journal of Protozoology*. **31**:233-240.
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PhD-related publications

7. Smith RD, Miranpuri GS, **Adams JH**, Ahrens E H (1985). *Borrelia theileri*: Isolation from ticks (*Boophilus microplus*) and tick-borne transmission between splenectomized calves. *American Journal of Veterinary Research*. **46**:1396-1398
 8. Smith RD, Levy MG, Kuhlenschmidt MS, **Adams JH**, Rzechula DL, Hardt TA, Kocan KM. (1986). Isolate of *Anaplasma marginale* not transmitted by ticks. *American Journal of Veterinary Research*. **47**:127-129.
 9. **Adams JH**, Smith RD, and Kuhlenschmidt, M.S. (1986). Identification of antigens of two isolates of *Anaplasma marginale*, using a western blot technique. *American Journal of Veterinary Research*. **47**:501-506.
 10. Pang VF, **Adams JH**, Beasley VR, Buck WB, Haschek WM. (1986). Myocardial and pancreatic lesions induced by T-2 toxin, a Trichothecene mycotoxin, in swine. *Veterinary Pathology*. **23**:310-319.
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Queensland Uni post doc publications

11. **Adams JH**, Monroy FG, East IJ, Dobson C. (1987). Surface and excretory/secretory antigens of *Nematospiroides dubius*. *Immunology and Cell Biology*. **65**:393-397.

12. **Adams JH**, East IJ, Monroy GF, Washington EA, Dobson C. (1987). Stage-specific antigens of *Nematospiroides dubius* Baylis, 1926 (Nematoda: Heligmosomoides). *Journal of Parasitology*. **73**:1164-1168.
13. **Adams JH**, Smith RD. (1988). Differential extraction of antigens of *Anaplasma marginale*. *American Journal of Veterinary Research*. **49**:257- 260.
14. **Adams JH**, Bushell G. (1988). The effect of protease inhibitors on *Eimeria vermiformis* invasion of cultured cells. *International Journal for Parasitology*. **18**:683-685.
15. **Adams JH**, East IJ, Monroy FG, Dobson C. (1988). Sex-specific antigens on the surface and in the secretions of *Nematospiroides dubius*. *International Journal for Parasitology*. **18**:999-1001.
16. **Adams JH**, Shiels IA, De Vos AJ (1989). Heterologous antibody responses of calves to *Anaplasma centrale* and *A. marginale*. *Veterinary Parasitology*. **31**:7-12.
17. Monroy FG, **Adams JH**, Dobson C, East IJ (1989). *Nematospiroides dubius*: influence of adjuvants on immunity in mice vaccinated with antigens isolated by affinity chromatography from adult worms. *Experimental Parasitology*. **68**:67-73.
18. Monroy FG, Dobson C, East IJ, **Adams JH**. (1989). Immunity in mice vaccinated with a molecular weight 60,000 glycoprotein secreted by adult *Nematospiroides dubius*. *International Journal for Parasitology*. **19**:71-76.
19. Monroy FG, **Adams JH**, Dobson C. (1989). Low molecular weight immunosuppressors secreted by adult *Nematospiroides dubius*. *International Journal for Parasitology*. **19**:125-127.
20. Monroy FG, Cayzer CJR, **Adams JH**, Dobson C. (1989). Proteolytic enzymes in excretory-secretory products from adult *Nematospiroides dubius*. *International Journal for Parasitology*. **19**:129- 131.
21. **Adams JH**, Bushell G. (1989). Changes in the cytoplasmic elements of cultured cells infected with *Eimeria vermiformis* sporozoites. *Journal of Protozoology*. **36**:131-136.
22. Monroy FG, **Adams JH**, East IJ, Dobson C. (1989). Excretory- secretory antigens from adult *Nematospiroides dubius*. *Immunology and Cell Biology*. **67**: 115-120.

NIH post doc publications

23. Good MF, Miller LH, Kumar S, Quakyi IA, Keister D, **Adams JH**, Moss B, Berzofsky JA, Carter R. (1988). A challenge for malaria vaccine development: limited immunological recognition of critical vaccine candidate antigens. *Science*. **242**:574-577.
24. Torii M, **Adams JH**, Miller LH, Aikawa M. (1989). Release of merozoite dense granules during erythrocyte invasion by *Plasmodium knowlesi*. *Infection and Immunity*. **57**:3230-3233.
25. **Adams JH**, Hudson DE, Torii M, Ward GE, Wellems TE, Aikawa M, Miller LH. (1990). The Duffy receptor family is located within the micronemes of invasive malaria merozoites. *Cell* . **63**:141-153. PMID:2170017.
26. Fang X, Kaslow DC, **Adams JH**, Miller LH. (1991). Cloning of the *Plasmodium vivax* Duffy receptor. *Molecular and Biochemical Parasitology*. **44**:125-132.
27. Dalton JP, Hudson D, **Adams JH**, Miller LH. (1991). Blocking of the receptor-mediated invasion of erythrocytes by *Plasmodium knowlesi* malaria with sulfated polysaccharides and glycosaminoglycans. *European Journal of Biochemistry*. **195**:789-794.

28. **Adams JH**, Sim BKL, Dolan SA, Fang X, Kaslow DC, Miller LH. (1992). A family of erythrocyte binding proteins in malaria parasites. *Proceedings of the National Academy of Sciences (USA)*. **89**: 7085-7089. PMID: PMC49650.
29. **Adams JH**, Fang X, Kaslow DC, Miller LH. (1992). Identification of a cryptic intron in the *Plasmodium vivax* Duffy binding protein gene. *Molecular and Biochemical Parasitology*. **56**:181-184.

Independent faculty career publications by year

30. Prickett MD, Smarz T, **Adams JH**. (1994). Dimorphism and intergenic recombination within the microneme protein (MP-1) gene family of *Plasmodium knowlesi*. *Molecular and Biochemical Parasitology*. **63**: 37-48.
 31. Tsuboi T, Kappe S, Al-Yaman F, Prickett MD, Alpers MP, **Adams JH**. (1994). Natural variation within the principal adhesion domain of the *Plasmodium vivax* Duffy binding protein. *Infection and Immunity*. **62**: 5581-5586. PMID: PMC303305.
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32. Kolakovich KA, Ssengoba A, Wojcik K, Tsuboi T, Al-Yaman F, Alpers MP, **Adams JH**. (1996). *Plasmodium vivax*: favored gene frequencies of the merozoite surface protein-1 and the multiplicity of infection of in a malaria endemic region. *Experimental Parasitology*. **83**: 11-18.
 33. Kappe SHI, **Adams JH**. (1996). Sequence analysis of the apical membrane antigen-1 (AMA-1) of *Plasmodium yoelii yoelii* and *Plasmodium berghei*. *Molecular and Biochemical Parasitology*. **78**: 279-283.
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34. Fraser T, Michon P, Barnwell JW, Noe AR, Al-Yaman F, Kaslow DC, **Adams JH**. (1997). Expression and serologic activity of a soluble recombinant *Plasmodium vivax* Duffy binding protein. *Infection and Immunity*. **65**: 2772-2777. PMID:9199499.
 35. Kappe SHI, Curley GP, Noe AR, Dalton JP, **Adams JH**. (1997). Erythrocyte binding protein homologues in rodent malaria parasites. *Molecular and Biochemical Parasitology*. **89**: 137-148.
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36. Kappe SHI, Noe AR, Blair PL, Fraser T, and **Adams JH**. (1998). A chimeric family of erythrocyte binding proteins of malaria parasites. *Proceedings of the National Academy of Sciences (USA)*. **95**: 1230-1235. PMID: PMC18728.
 37. Michon P, Fraser T, Herrera M, Herrera S, **Adams J**. (1998). Serologic responses to the *Plasmodium vivax* Duffy binding protein in a Colombian village. *The American Journal of Tropical Medicine and Hygiene*. **59**:597-599.
 38. Noe AR, **Adams JH**. (1998). *Plasmodium yoelii* YM MAEBL protein is coexpressed and colocalizes with rhoptry proteins. *Molecular and Biochemical Parasitology*. **96**:27-35.
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39. Noe AR, Fishkind DJ, **Adams JH**. (2000). Spatial and Temporal Dynamics of the Secretory Pathway during Differentiation of the *Plasmodium yoelii* Schizont. *Molecular and Biochemical Parasitology*. **108**: 169-185.
 40. Michon P, Fraser T, **Adams J**. (2000). Naturally acquired and vaccine elicited antibodies block erythrocyte cytoadherence of the *Plasmodium vivax* Duffy binding protein. *Infection and Immunity*. **68**: 3164-3171. PMID: PMC97553.

41. Xainli J, **Adams JH**, King CL. (2000). The Erythrocyte Binding Motif of *Plasmodium vivax* Duffy Binding Protein is Highly Polymorphic and Functionally Conserved in Isolates from Papua New Guinea. *Molecular and Biochemical Parasitology*, 111: 253-260. PMID:11163434.

42. Shirano M, Tsuboi T, Kaneko O, Tachibana M, **Adams JH.**, Torii M. (2001). Conserved Regions Revealed by the Comparison of *Plasmodium yoelii* 100-kDa Rhoptry Protein with *P. falciparum* Homologue. *Molecular and Biochemical Parasitology*, 112: 297-299.
43. Michon P, Wooley I, Wood E, Kastens W, Zimmerman PA, **Adams JH.** (2001). Duffy-null promoter heterozygosity reduces DARC expression and abrogates adhesion of the *P. vivax* ligand required for blood-stage infection. *FEBS Letters*, 495:111-114.
44. **Adams JH**, Kaneko O, Peterson DS, Blair PL. (2001). An Expanding *eb1* Family of *Plasmodium falciparum*. *Trends in Parasitology*, 17:297-299.
45. Kappe SHI, Gardner MJ, Brown SM, Ross J, Matuischewski K, Ribiero JM, **Adams JH**, Carucci DJ, Hoffman SL, Nussenzweig V. (2001). Exploring the transcriptome of the malaria sporozoite stage. *Proceedings of the National Academy of Sciences (USA)*, 98, 9895-9900.
46. Fraser T, Kappe SHI, Narum DL, VanBuskirk KM, **Adams JH.** (2001). Identification of Erythrocyte Binding Activity by Selected Domains of the Malaria Vaccine Candidate Apical Membrane Antigen-1 As Expressed on the Surface of Transfected COS-7. *Molecular and Biochemical Parasitology*, 117, 49-59.

47. Blair PL, Witney A, Moch JK, Haynes JD, Carucci DJ, **Adams JH.** (2002). Quantification of developmentally regulated gene transcripts in *Plasmodium falciparum* using real-time RT-PCR. *Nucleic Acids Research*, 30(10), 2224-2231.
48. Blair PL, Kappe SHI, Maciel J, Balu B, **Adams JH.** (2002). *Plasmodium falciparum* *maebl* is a unique member of the *eb1* family. *Molecular and Biochemical Parasitology*, 122,35-44.
49. Michon P, Stevens JR, Kaneko O, **Adams JH.** (2002). Evolutionary relationships of conserved cysteine-rich motifs in adhesive molecules of malaria parasites. *Molecular Biology and Evolution*, 19, 1128-42. PMID:12082132.
50. Cole-Tobian JL, Cortes A, Baisor M, Kastens W, Xainli J, Bockarie M, **Adams JH**, King CL. (2002). Age-acquired immunity to a *Plasmodium vivax* invasion ligand, the Duffy binding protein. *Journal of Infectious Diseases*, 186(4), 531-9. PMID:12195381.
51. Xainli J, Baisor M, Kastens W, Bockarie M, **Adams JH**, King CL. (2002) Age-dependent cellular immune responses to *Plasmodium vivax* Duffy binding protein in humans. *Journal of Immunology*, 169, 3200-3207. PMID:12218138.

52. Balu B, **Adams JH.** (2003). Fluorescent chloramphenicol as a substitute for radioactive [¹⁴C]-chloramphenicol for CAT reporter assays in *Plasmodium falciparum*. *Molecular and Biochemical Parasitology*, 126, 285-286.
53. Xainli J, Cole-Tobian JL, Baisor M, Kastens W, Bockarie M, Yazdani SS, Chitnis CE, **Adams JH**, King CL. (2003). Epitope-Specific Humoral Immunity to *Plasmodium vivax* Duffy Binding Protein. *Infection and Immunity*, 71, 2508-2515.

54. Ohas EA, **Adams JH**, Waitumbi JN, Orago ASS, Barbosa A, Lanar DE, Stoute JA. (2004). Measurement of antibody levels against region II of the erythrocyte binding antigen 175

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(Mentees noted since 2015 for my USF senior author publications or research initiated in my group: postdoctoral fellows/Research Associate; PhD student; other student trainees)

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